Amiprox

Product Information Sheet

ProtiPlate Pop2

Depletion of
Human Serum Albumin (HSA)&
Immunoglobulin G (IgG)

96-well deepwell filterplate

Storage Temperature: 2 - 8°C

Product: AMI002HSA-IgG

Do not freeze.



Product Description

Plasma is one of the most information-rich yet analytically complex biological fluids due to its extraordinary dynamic range of protein concentration. Highly abundant proteins like human serum albumin (HSA) and immunoglobulins (IgG) dominate the proteome, accounting for nearly 70 % of the total protein mass, while clinically relevant low-abundance proteins may be present at concentrations up to 12 orders of magnitude lower. o overcome this challenge, ProtiPrep™ Pop2 utilizes a capture system combining specific ligands for Human Serum Albumin (HSA) and Protein A for Immunoglobulin G (IgG) capture, depleting >97% of HSA and IgG from 10 µl of plasma within 10 minutes with minimal nonspecific interaction.

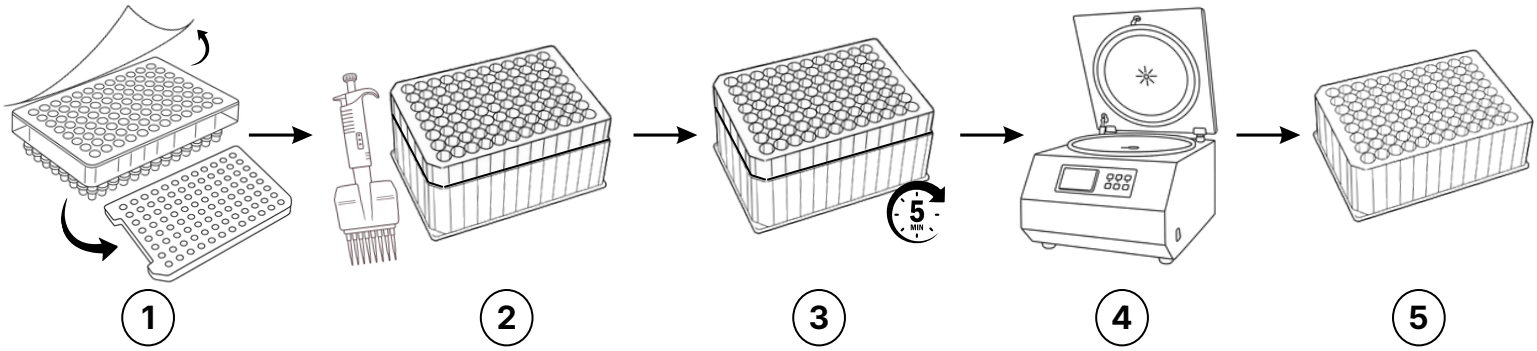
Additional Information

ProtiPlate Pop2 deepwell filterplates are pre-packed with the optimum amounts of functionalized agarose beads (155 µl settled beads) and PBS buffer (pH 7.4, 0.02 % sodium azide), to ensure efficient and reproducible sample preparation without the need for special laboratory equipment (Centrifuge capable of operating 1000 x g).

Kit Contents:
Included: pre-packed filter plate (resin + buffer), collection plate, foil seal, silicone mat.
Not provided: microcentrifuge (1000 x g), pipette/tips, EDTA plasma sample.
Validated for human EDTA plasma only; not tested with other anticoagulants, serum, or species.
Note: No extra reagents or solvent exchange required.
Note: Single use only; do not reuse resin.
For Research Use Only.

Handling

ProtiPlate Pop2 simplifies sample preparation to the maximum. Apply 10 µl of undiluted plasma (use filter tips) to each well, mix by pipetting up and down (use wide bore tips), incubate, centrifuge, and collect the sample. The prepared sample (~150 µl) is stored in PBS buffer (pH 7.4, 0.02 % sodium azide) and ready for analysis or subsequent processing. However, the amount of HSA and IgG in plasma can vary considerably. For best results, optimize the ratio of sample to slurry volume for each specific application.



Step 1: Equilibrate the ProtiPlate Pop2 to room temperature. Peel off the aluminium foil and remove the silicon mat from the bottom of the plate. Place the filter plate on top of the collection plate.

Step 2: Add 10 µL of undiluted plasma to each well. Pipette the slurry in each well at least 10 times up and down to ensure proper mixture of the sample with the resin.

Step 3: Incubate at room temperature for 5 minutes. Alternatively, place the plate on an orbital shaker and incubate for 5 minutes.

Step 4: Centrifuge for 2 minutes at 1000 x g.

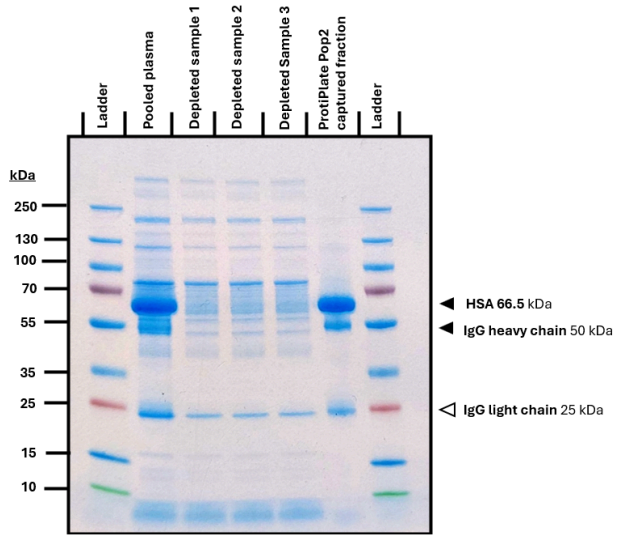
Step 5: Discard the filter plate. The HSA/IgG depleted samples remain in the collection plate. The samples may be used immediately for further processing or stored at –20 °C for later use.

Optional: To maximize sample recovery, add your desired volume of PBS directly to each well and centrifuge again for 2 minutes at 1000 x g. The additional eluate will be collected in the same collection plate. Note: the volume of PBS added will equal the additional volume recovered, so adjust accordingly based on your downstream application.

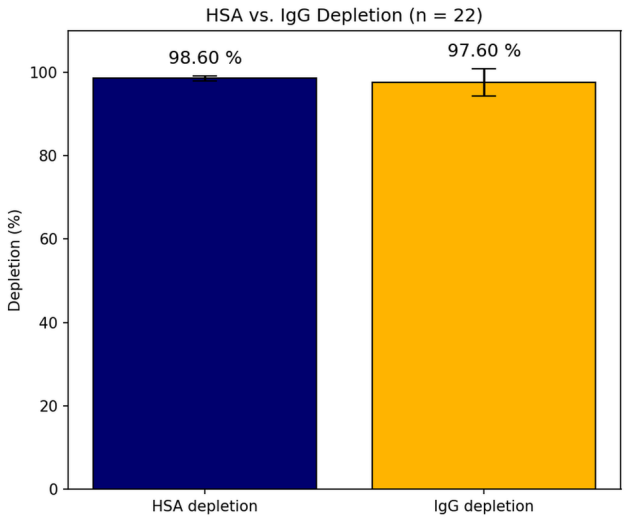
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Depletion Efficiency



SDS-PAGE analysis of pooled human plasma (60 donors) before and after HSA and IgG depletion using ProtiPlate Pop2. Depleted samples show strong reduction of HSA and IgG bands, while the bead fraction confirms specific capture.



ELISA-based depletion performance of ProtiPlate Pop2 was evaluated using pooled human plasma (60 donors, n = 22). Mean depletion efficiencies of 98,60 % (HSA) and 97,60 % (IgG) were observed, with median efficiencies of 98,75 % and 98,23 %, respectively.

Note: The data shown correspond to processing of 10 µL pooled human plasma (~700 µg total protein) per column.

Note: Sample processing will depend on the type of downstream analysis and may require buffer exchange, lipid and other metabolite removal and/or concentration for 2D gel electrophoresis or mass spectrometry analysis after depletion.

Troubleshooting

Observation	Possible cause	Recommended action
HSA and IgG were not completely removed.	Sample exceeded binding capacity.	Reduce the amount of sample processed.
	Incomplete binding.	Increase the incubation time.
	Sample was not mixed during incubation.	Mix the sample with resin gently 10 times.
HSA and IgG were not completely removed.	Centrifuge did not reach the required 1000 x g, or the foil seal and silicon mat were not fully removed in Step 1.	Confirm the centrifuge reaches 1000 x g for the full 2 minutes, and verify both seals were removed before loading.
	Filter membrane is damaged or compromised.	Inspect the plate before use. If damage is visible on receipt, contact technical support before proceeding.
	Resin settled unevenly during shipping or storage, or mixing was inconsistent between wells.	Gently tap or invert the sealed plate to resuspend the resin before removing the foil, and ensure every well receives the full 10 times up and down mixing.
Resin appears dry or caked on arrival.	Temperature excursion during shipping, or a compromised foil seal.	Do not use the plate. Contact technical support and your quality team before proceeding.

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